

This short presentation shows the results of three unprecedented experiments possible only with the **Olis CLARiTY 1000**.

The photo shows the novel DSPC within the dual beam sample compartment.



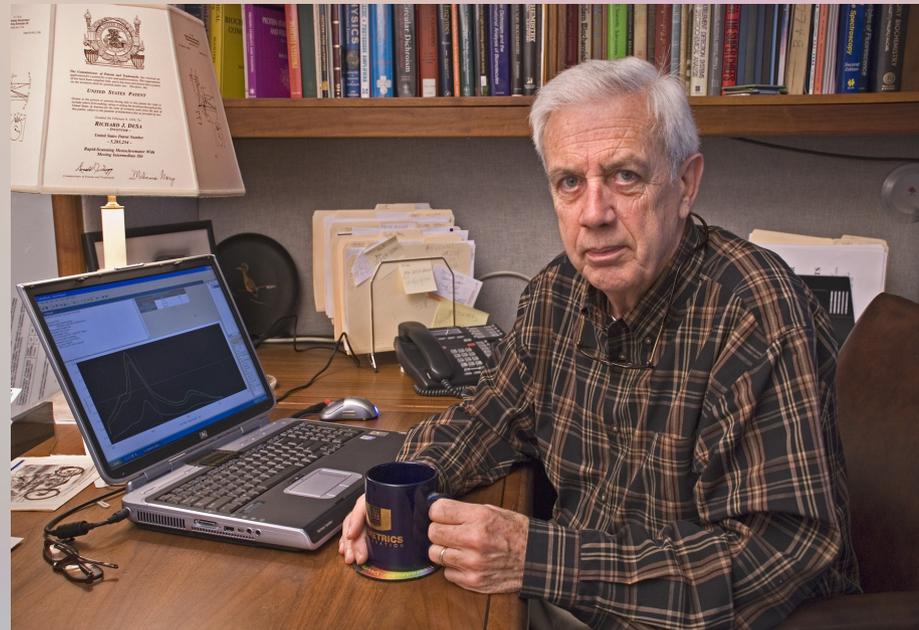
**An entirely new era  
of studying scattering biological suspensions  
has been entered.**

You have gone full circle, from the reversion spectroscope at the start of the 20th century to the detector that we all want in the 21st century.

That is truly cool. Best Wishes – John S. Olson  
Ralph and Dorothy Looney Professor of Biochemistry and Cell Biology  
Rice University

Personal correspondence, July 2010

The three experiments were conducted by Dr. Richard DeSa –  
the inventor of both the Rapid-Scanning Monochromator with Moving Intermediate Slit  
(aka, Olis RSM 1000) that he used to collect  
the following 300 nm width scans at a rate of 2 per second  
and  
the DeSa Suspension Presentation Cavity (DSPC, seen on the first page  
and key to the CLARiTY performance) –  
using an exceedingly dilute suspension of intact human red blood cells.



## Experiment 1:

Watching as intact live red blood cells change from oxygenated to deoxygenated as the environmental O<sub>2</sub> is consumed.

Figure 1: The spectrophotometric recording of the release of O<sub>2</sub> from HbO<sub>2</sub> in human red blood cells (RBC).

300 nm span scans, collected at a rate of 2/second for 220 seconds.

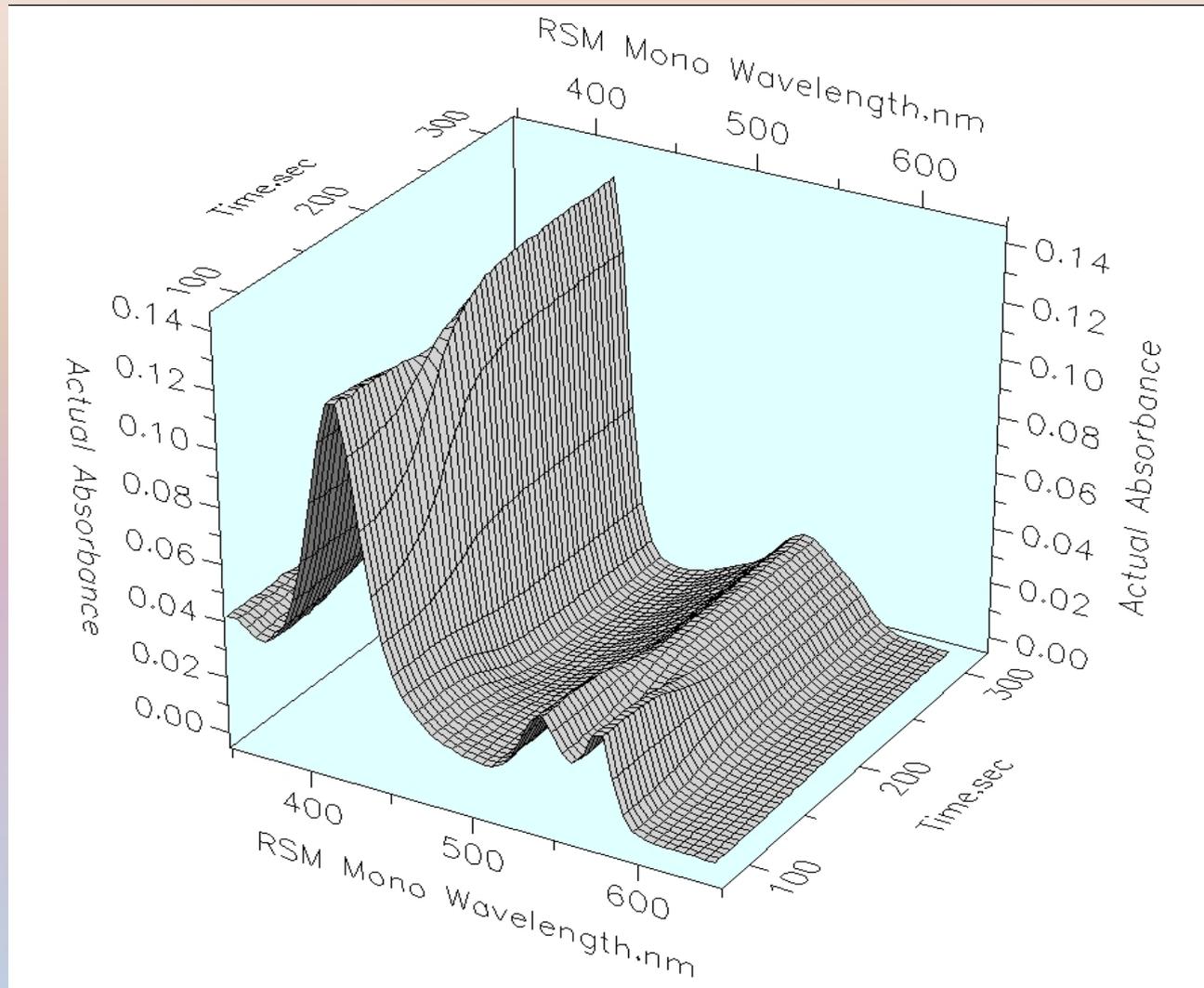
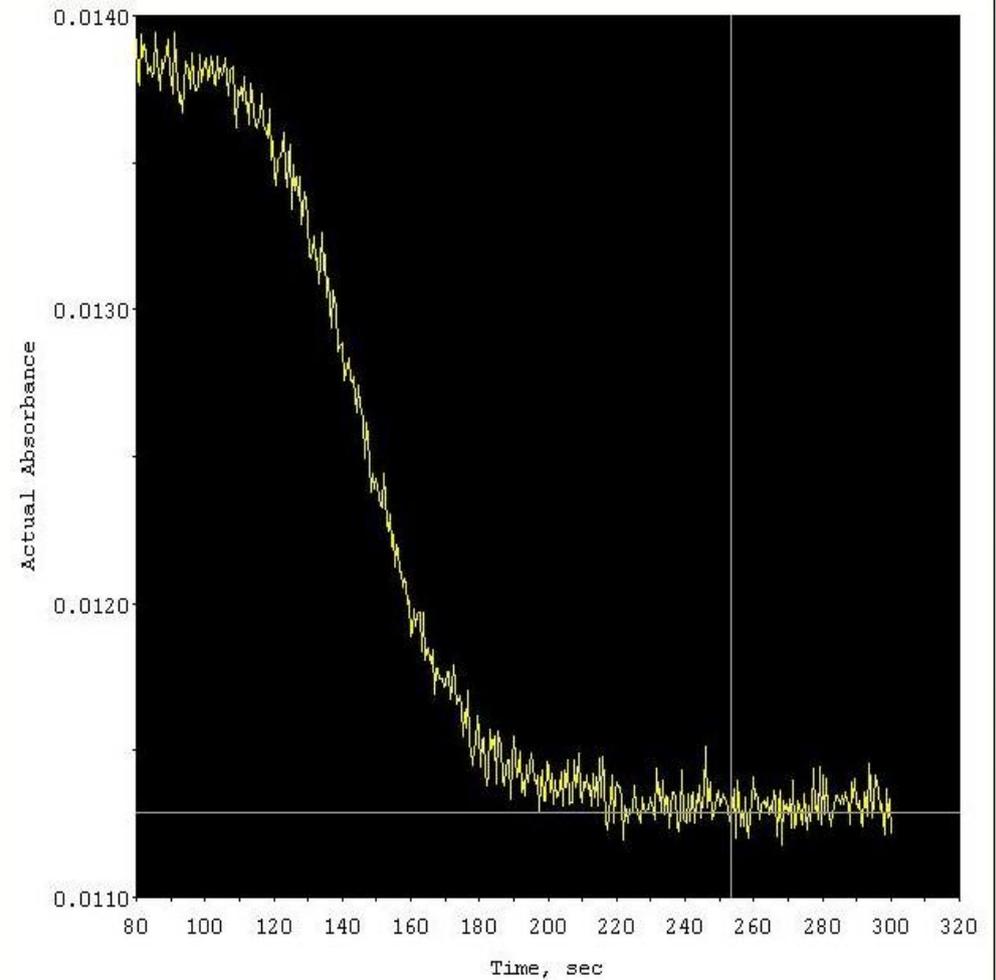
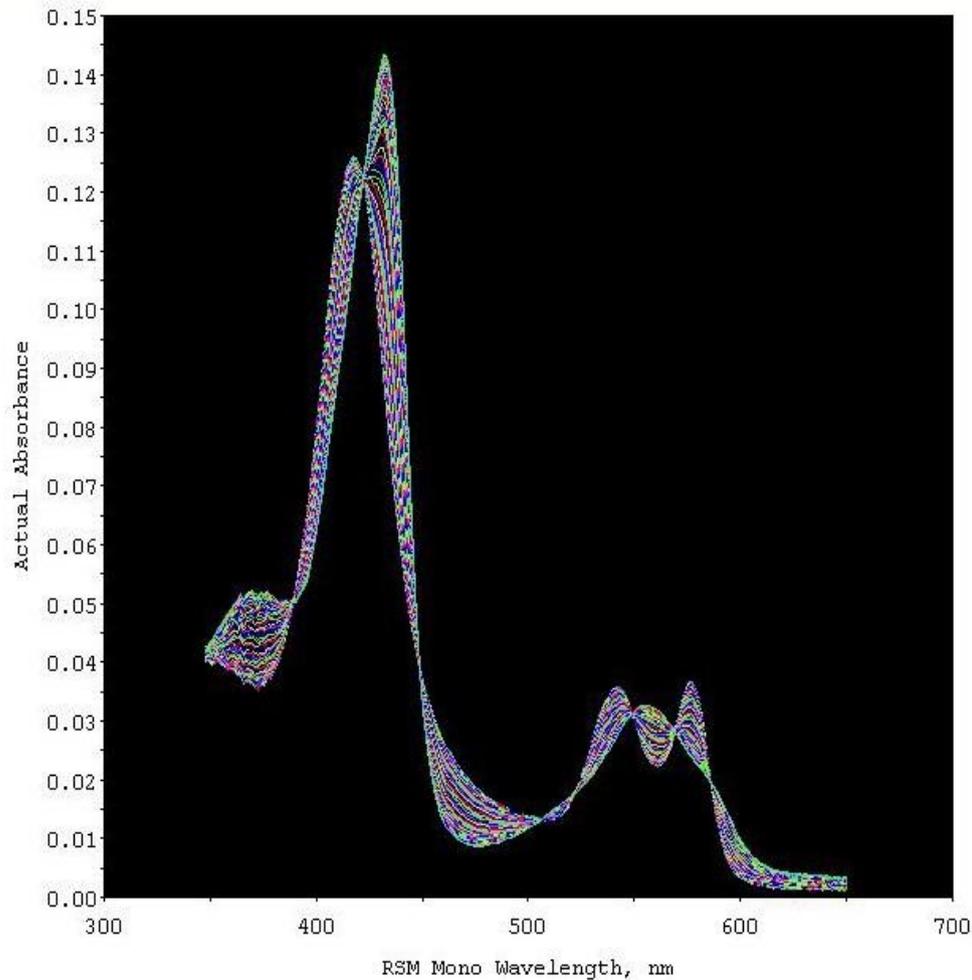


Figure 2 Clarifies what was shown in the 3D image. The graph to the right shows the kinetics.

The cells started oxygenated, the downward sloping line is the absorbance at 500 nm as the oxygen decreases, and the flat line at the bottom tells us that all of the HbO<sub>2</sub> has released its oxygen.

**Spectra (300 nm width @ 2 scans/ second)**

**Abs change at 500 nm (notice Y axis!)**



***No other spectrophotometer has ever  
collected spectra of such quality  
from intact RBCs in suspension.***

***Because ...***

**RBC suspensions scatter the light so much that conventional spectrophotometers are unable to collect accurate spectra at any rate (let alone at 2/sec).**

*The total immunity of the CLARiTY to scatter,*

**its high photometric accuracy,**

**and**

**the high speed wavelength scanning of the RSM  
combine**

to **allow the study and precise mathematical analysis**  
of real-time high speed functioning of  
the HbO<sub>2</sub> system.

**This instrument is *ideal* for advancing  
our understanding of this centrally  
important function in animal life.**

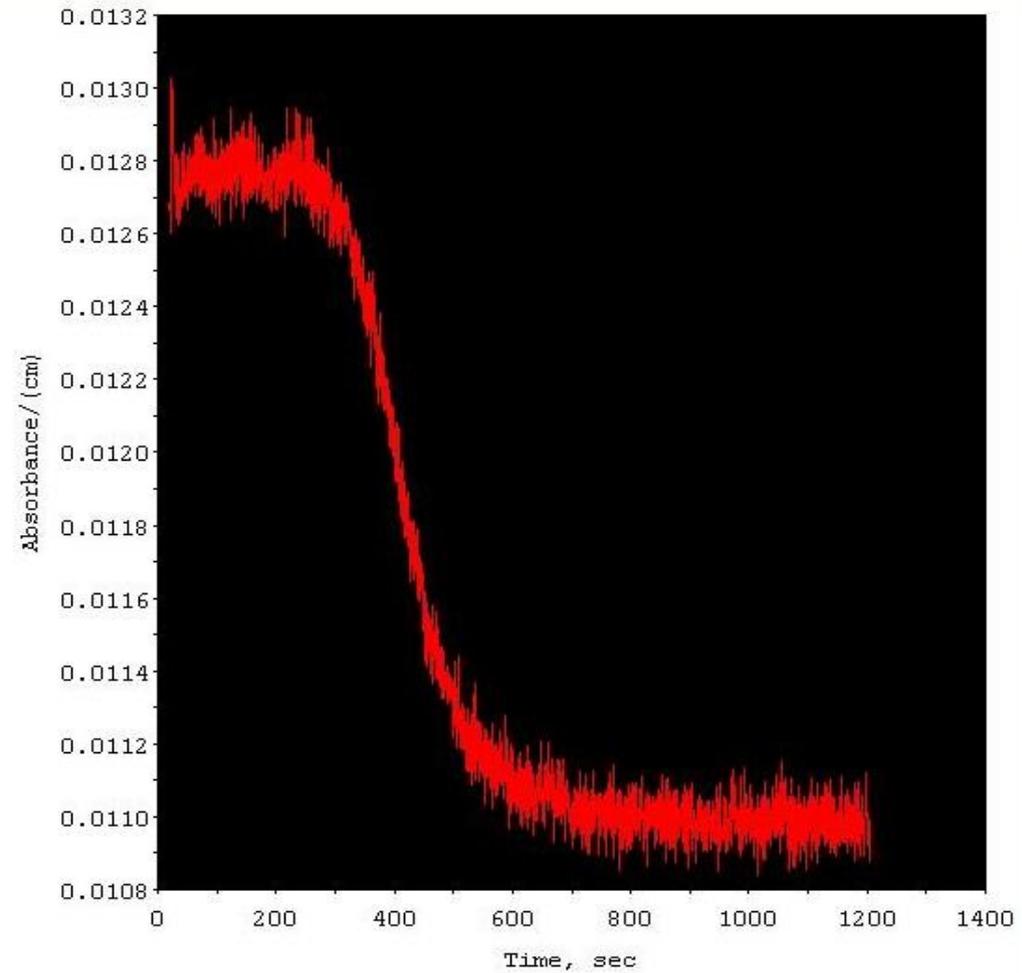
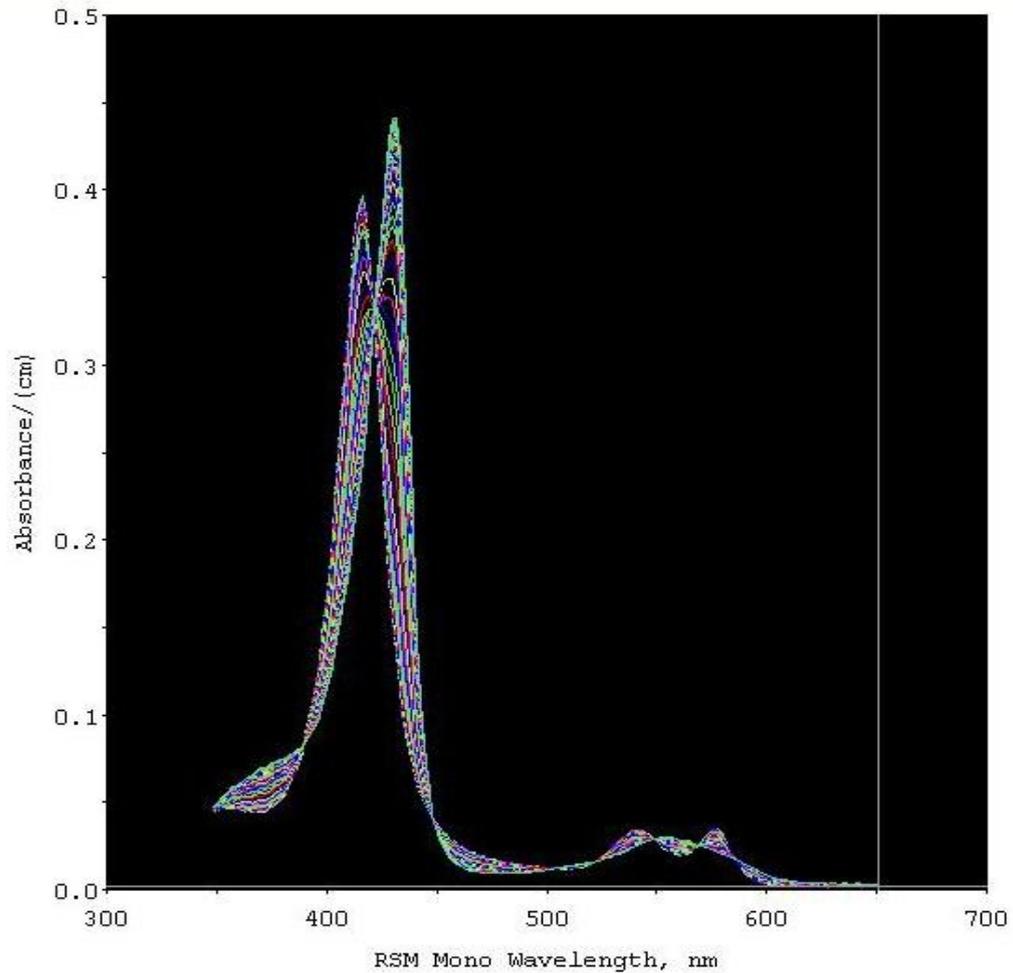
It is *only* with this technology that one can truly answer the question  
“**How does Hb function in real life?**”

## **Experiment 2: Deoxygenation of Hb released from lysed RBCs.**

***This is the condition under which most experiments of HbO<sub>2</sub> are done, because intact RBC cannot be studied in conventional spectrophotometers (of the last 150 years)***

- RBCs were put in the DSPC in phosphate buffer without NaCl.
- The cells lysed in less than 20 seconds and the data shown were collected.
- There is a substantial and meaningful increase in the absorbance of the Soret region since the packing effect (hypochromism) seen with intact RBCs no longer is present.
- The deoxygenation occurs with no apparent shift in the isobestic point as the HbO<sub>2</sub> changes to Hb.

This is similar to the result obtained when intact RBCs are examined  
(i.e, data shown in figures 1 and 2).



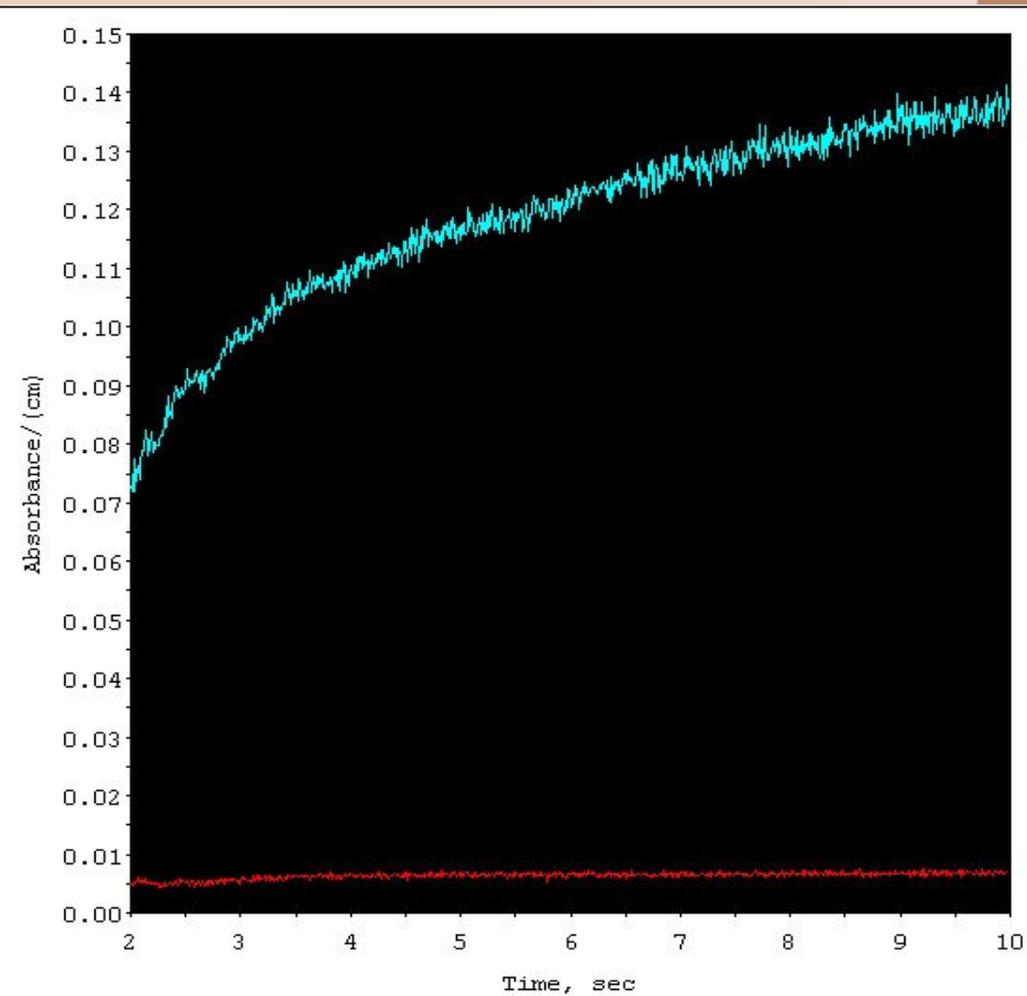
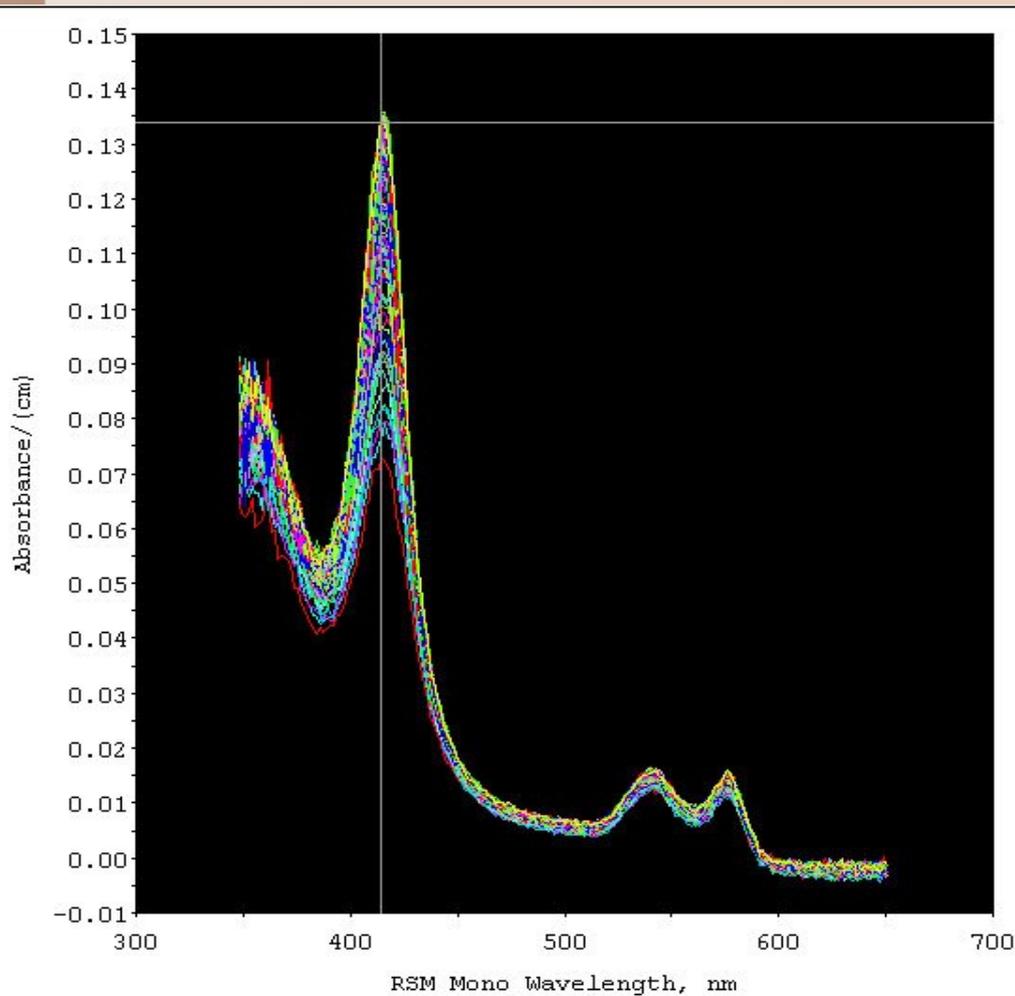
**“The Olis CLARiTY offers the simplest and most direct way of studying a bio-molecule under these two conditions, oxygenated hemoglobin tightly packed in the RBC or free in solution.”**

**Lawrence J. Parkhurst, Ph.D.**  
Hewett University Professor, UN Lincoln

(whose blood cells were used for the experiments!)

## Experiment 3: *The process of the HbO<sub>2</sub> releasing from the RBCs as they lysed during a 10 second time period.*

The data transition from very good spectra of HbO<sub>2</sub> in intact RBC (Figure 1) to very good spectra of free HbO<sub>2</sub> in a clear buffer solution (Figure 3).



**Today's three experiments prove that you can  
now do spectrophotometry on highly  
scattering suspensions.**

Imagine loading your Olis CLARiTY's DSPC

*with suspensions*

*(mitochondria, yeast, bacteria, algae, nanoparticles ...)*

and getting accurate absorbance spectra,\*

(\*And, with the right additional hardware, fluorescence spectra, too!)

# The Olis CLARiTY

- **Total immunity to the effects of scatter**
- Cuvettes with effective pathlengths exceeding 20 cm, providing **dramatic sensitivity enhancement**
- High speed wavelength scanning (100 scans/ second), so that **full spectra are available to follow kinetics**
- Dual beam optical system with photomultiplier tube detectors for **best photometric precision and accuracy**

“Not a comment but an opinion -

This system is an almost incredible advance on the best  
Britton Chance was ever able to do for turbid suspensions.  
I do hope OLIS is able to sell a lot of them.”

Quentin H. Gibson  
Emeritus Prof, Cornell Univ  
Personal correspondence, July 2010

The spectra were really beautiful and impressive . . . Very exciting, and beautiful results!

Larry J. Parkhurst

The CLARiTY changes everything with respect to thinking in spectrophotometry. Awesome! I just have to have one in my lab. Will be writing grants later this year to include a purchase.

Paul S. Hoffman  
Professor, University of Virginia School of Medicine

**Clear, unambiguous spectra  
at rates adjustable to the system under study.**

**What research projects can you pursue now since  
Scatter Does Not Matter?**

**Get more information on the Olis CLARiTY spectrophotometers  
by calling **1.706.353.6547** or  
writing to **sales@olisweb.com****

*If he's available, Dr. DeSa himself will be happy to speak with you ,too!*